Biomolecular Films for Direct Air Capture of CO₂

BETO Peer Review April 3, 2023 | Denver, CO Karsten Zengler, Juan Tibocha-Bonilla, Martin Gross, Drew Greene, Ashton Zeller, Luke Dahlin, Matt Wiatrowski, Ryan Davis, Mike Guarnieri







Background: CO₂ Is A Key Cost Driver in Algal Biofuels



- CO₂ accounts for ~20% of total costs of Algal Biomass Selling Price (ABSP).
- ABSP is a function of productivity.
- Technologies that enable direct air capture (DAC) of atmospheric CO₂ to decouple algae cultivation from CO₂ point sources and enhance productivity present an opportunity to improve the economics and resource potential of algal biomass.

FY20 Bioenergy Technologies Multi-Topic FOA

Topic Area 3 Metrics:

The application must propose to meet all the minimum targets in the table below by the end of the project.

DE-FOA-0002203 Topic Area 3: Algae Bioproducts and CO2 Direct-Air-Capture Efficiency (ABCDE)

- FOA Issue Date: 1/23/2020
- Award Start Date: 7/31/2020
- Initial Verification: 12/1/2020

Metrics	Unit	Minimum	Stretch	
Algal biomass revenue potential	\$ per ton harvested algae biomass	25% increase from applicant's baseline*	50% increase	
Algal biomass quality for downstream testing	% meeting fuel and product(s) specifications	<10% out of specification	<5% out of specification	
Algae areal productivity	g/m²/d	Increase productivity 10% over applicant's baseline with CO ₂ from DAC	Increase productivity >10% over applicant's baseline with CO ₂ from DAC	
DAC CO ₂ delivered and utilized by algal system	% of DAC CO ₂ delivered and utilized by algal system	20% increase over applicant's baseline	>20% increase over applicant's baseline	
Cost** of CO ₂ delivered to algal system	\$ per volume of CO ₂ delivered to the algae system from DAC versus non-DAC	10% decrease in the cost of CO ₂ delivered via DAC versus non- DAC CO ₂ delivery	>10% decrease in cost of CO ₂ delivered via DAC versus non- DAC CO ₂ delivery 3	

Project Overview

Objective: We propose to integrate recent advances in computational metabolic modeling, algal genetic engineering, algal cultivation, and algal biomass upgrading to enable secretion of carbonic anhydrase for enhanced CO_2 capture and conversion in immobilized algal biofilms.

End-Project Goal: Achieve a 25% increase in algal biomass revenue potential, algal biomass quality <10% out of specification for downstream testing, a 10% increase in productivity, a 20% increase in CO_2 obtained from ambient air, and a 10% decrease in CO_2 costs via bio-based air capture technology.

Approach: Team and Task Structure

UC San Diego

Genome-scale Metabolic Model, Engineering Designs Task Lead: Karsten Zengler



Enzyme and Strain Engineering | Task Lead: Mike Guarnieri TEA/LCA | Task Leads: Ryan Davis and Matt Wiatrowski



Revolving Algal Biofilm (RAB) System Deployment Task Lead: Martin Gross



Product Development | Task Lead: Ashton Zeller

Approach: Task 2 - Computational Modeling

Task 2 will reconstruct a metabolic model of *Picochlorum renovo* and deploy this model for the design of improved protein production and secretion to achieve maximal productivity in this microalga.

- Sub-task 2.1: Genome-scale metabolic model reconstruction.
- Subtask 2.2: Model Refinement and Rational Strain Engineering Design



Milestone 2.1.1 (M6): Reconstruction of a genome-scale metabolic model of *P. renovo*.

Milestone 2.2.1 (M18): GEM Refinement: integrate phenotypic and omic data and all possible secretion and carbonic anhydrase (CA) sequences into the model and predict optimized secretion without loss in productivity.

Approach: Task 3 - CA Production and Algal Strain Engineering





Task 3 will target expression and down-selection of candidate CA variants. Top-candidate CA will be incorporated into *P. renovo* to enable photoproduction and secretion. This work will ultimately deliver i) CA variants suitable for cultivation supplementation and ii) algal biocatalysts with *in situ* CA photoproduction capacity.

- Sub-task 3.1: CA Production
- Sub-task 3.2: Algal Strain Engineering

Milestone 3.1.1 (M15): Down-select and produce >10g of two top-candidate CA in *P. pastoris* for delivery to GWT for assessment in RAB systems.

Milestone: 3.2.1 (M30): Demonstrate functional extracellular secretion of a heterologous CA. SDS-PAGE, Western Blot, and/or densitometric analyses will be employed to quantify protein in the culture secretome.

Approach: Task 4 – RAB Deployment & Compositional Analysis



Task 4 will assess algal biomass yield and productivity increases as a function of CA supplementation and/or algal secretion to further inform CA down-selection, define CA concentration requirements, and assess process enhancement metrics.

Milestone 4.1 (M9): Establish a productivity baseline, reporting g/m²/day ash-free dry weight, and composition for *P. renovo* in a lab-scale RAB system.

Milestone 4.2 (M21): Deploy top-candidate exogenously supplemented CA (2) in >100L RAB systems to establish algal productivity and conversion yield enhancement relative to baseline, reporting biomass productivity and AFDW composition.

Milestone 4.2 (M33): Deploy engineered algal CA secretion strains at 800L scale, with semi-continuous harvest, reporting biomass productivity and AFDW composition.⁸

Approach: Task 5 - Product Development



Task 5 will focus on the development of a novel thermoplastic composite using RAB-derived algae produced in Task 4.

Milestone 5.1 (M12): Compositional Analysis for Bioplastic Conversion. RAB biomass compositional analysis from Task 4 will be evaluated to determine expected performance in bioplastic conversion.

Milestone 5.2 (M27): Formula Development and Compounding. Develop a formula and compound it into a 45% algae masterbatch suitable for injection molding test parts.

Milestone 5.3 (M35): Injection Molding and Material Property Testing. Injection mold parts containing 20% algae and then test them for material properties such as MFI, moisture, tensile, flexural, and other characteristics.

Approach - Task 6: TEA/LCA

Task 6 will serve to assess mass and energy balances in the proposed CA-enhanced deployment process, ultimately defining an economically-viable and sustainable path to commercialization via iterative TEA/LCA model establishment and refinement.



Milestone 6.1 (M24): Modification of the established CAP TEA model with modeled carbon utilization efficiency enhancements. Establish LCA model for utilization of atmospheric CO₂ enabled by CA supplementation and/or secretion. **Milestone 6.3 (M36):** Refine models with BP2 inputs and deliver techno-economic path to commercialization.

Progress and Outcomes

Mathematical model of Picochlorum renovo

*i*CZ1179-Picre → 6,374 reactions, 3,587 metabolites, and 1,179 genes

Mathematical model of Picochlorum renovo

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Carbon source	Dark or Light	Experimental	Simulation	Conclusion
glycerol	Dark	Non-Growth	Non-Growth	TN
arabinose	Dark	Non-Growth	Non-Growth	TN
acetate	Dark	Non-Growth	Non-Growth	TN
galactose	Dark	Non-Growth	Non-Growth	TN
formate	Dark	Non-Growth	Non-Growth	TN
pyruvate	Dark	Non-Growth	Non-Growth	TN
glucose	Dark	Non-Growth	Non-Growth	TN
glutamine	Dark	Non-Growth	Non-Growth	TN
uridine	Dark	Non-Growth	Non-Growth	TN
fumarate	Dark	Non-Growth	Non-Growth	TN
succinate	Dark	Non-Growth	Non-Growth	TN
sorbitol	Dark	Non-Growth	Non-Growth	TN
lactate	Dark	Non-Growth	Non-Growth	TN
ribose	Dark	Non-Growth	Non-Growth	TN
glycerol	Light	Non-Growth	Non-Growth	TN
arabinose	Light	Non-Growth	Non-Growth	TN
acetate	Light	Growth	Growth	ТР
galactose	Light	Non-Growth	Non-Growth	TN
formate	Light	Non-Growth	Non-Growth	TN
pyruvate	Light	Growth	Growth	ТР
glucose	Light	Non-Growth	Non-Growth	TN
glutamine	Light	Growth	Growth	ТР
uridine	Light	Growth	Growth	TP
fumarate	Light	Non-Growth	Non-Growth	TN
succinate	Light	Non-Growth	Non-Growth	TN
sorbitol	Light	Non-Growth	Non-Growth	TN
lactate	Light	Non-Growth	Non-Growth	TN
ribose	Light	Growth	Growth	ТР

True Positives	5
True Negatives	23
False Positives	0
False Negatives	0

100%
100%
100%

Mathematical model of Picochlorum renovo

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Growth under light conditions

Using NH₄ as nitrogen source

- Acetate
- Pyruvate
- Glutamine
- D-Ribose
- Uridine



Protein Secretion Network

- We have successfully finalized the main requirement to build a secretory model: The Protein-Specific Information Matrix (PSIM)
- 687 (out of 2071) proteins were identified to contain a Signal Peptide
- PSIM was constructed from homology with *Chlamydomonas reinhardtii* and from available information in UniProt database



Heterologous Expression of CA

Expression cassettes Tf binding site Α В С D ble mCherry = SP P_{FIf} $P_{Rubisco}$ Tf ble CA P_{Flf} P_{Rubiscc}

- Successful CA expression and secretion achieved in *P. pastoris*.
- Over 12 unique CA were evaluated for secretion potential and downselected based upon solubility, stability, and CA activity in *P. renovo* cultivation media.

CA Supplementation Enhances Growth



Enhanced Secretion Achieved in P. renovo

Establishment of a functional synthetic transcription factor in P. renovo



P. renovo lab-scale RAB trial results

■ Lipid ■ Carb ■ Protein



- > 25g/m²/day productivity in RAB configuration under ambient CO₂
- > 50g/m²/day productivity in RAB configuration under ambient CO₂ with CA supplementation
- RAB biomass composition has minimal differential relative to pond cultivated biomass





Plasticization Trials Reflect Favorable Biomass Composition

- Algix conducted compositional testing and showed that RAB biomass is suitable for plastic composite production.
- The algae contained only 6 compounds associated with degradation in the odor testing.
 - This is below current thresholds for deeming algae biomass significantly degraded and qualifies the RAB algae material for use in commercial applications.
- Protein, ash, and water contents met minimum requirements.







Modified Plasticization

(Go/No-Go): Year 2 Q7 – Interim Validation Period. Demonstrate a 10% increase in algal biomass revenue potential, algal biomass quality <15% out of specification for downstream testing, a 5% increase in productivity, a 10% increase in CO_2 obtained from ambient air, and a 5% decrease in CO_2 costs via enhanced air capture.



Preliminary results (RAB data extrapolated to open ponds)

- CA productivity increase:
 - Revenue +20%
 - MFSP -10%
 - *25.3 → 51.7 g/m²/day
- CO₂ from Ambient capture:
 - 18% MFSP decrease
- Fuel Yield: 54 \rightarrow 71 GGE/ton
- Solid coproduct
 - Revenue up to +27%
- Further improvements possible with integration of RAB system (in progress)

(Go/No-Go): Year 2 Q7 – Interim Validation Period. Evaluate Hy-CA pond supplementation at 100mL outdoor simulation scale and >100L RAB scale, demonstrating a 10% increase in algal biomass revenue potential, algal biomass quality <15% out of specification for downstream testing, a 5% increase in productivity, a 10% increase in CO_2 obtained from ambient air, and a 5% decrease in CO_2 costs via enhanced air capture.

■ Lipid ■ Carb ■ Protein

Target Protein Content: 53.5% ± 15% **Achieved Content:** 51% ± 11%



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- All Go criteria successfully achieved.
- BP2 Completed in October, 2022
- BP3 Initiated in January, 2023
- BP3 will focus upon:
 - GEM optimization using multi-omic and physiological inputs
 - Secretion optimization using GEM-informed strain engineering designs
 - Scale-up to 800L RAB system
 - CAP processing of RAB biomass and conduct injection molding
 - Refinement of TEA/LCA models

Impact

- Moving the needle on algae productivity: achieved >50g/m²/day areal productivity from atmospheric CO₂.
- Enhances algal biomass revenue potential via reduction (elimination) of CO₂ sparging requirements, improved bioproductivity, and high-value co-production.
- Established comprehensive *Picochlorum* genome-scale model for basic and applied research pursuits.
- RAB cultivation enables in-line harvesting with dramatic reduction in dewatering requirements.
- 30% fuel yield improvement and nearly 20% MFSP improvement relative to baseline.
- Carbon intensity reduction achieved via improved carbon utilization efficiency and green biopolymer production.
- Bio-based DAC has a number of advantages over abiotic DAC technologies including minimal CAPEX/OPEX relative to current and emerging abiotic CO₂ capture and delivery systems, which require substantial water and energy inputs²⁶

Summary

- **Problem:** CO₂ accounts for nearly 20% of ABSP
- **Solution:** We are pursuing and integrated computational modeling, strain engineering, cultivation engineering, and biomass valorization strategy to enable high-productivity biomass with dramatically reduced CO₂ requirements.
- Progress to Date
 - Established a complete genome-scale metabolic model for the high productivity microalga, *P. renovo*.
 - Successfully engineered *P. renovo*, for secretion of carbonic anhydrase.
 - Deployed *P. renovo* in a RAB configuration, achieving > 50g/m²/day biomass productivity on atmospheric CO₂.
 - Initiated co-production of biopolymer(s) from RAB-derived biomass
 - Generated preliminary TEA/LCA models for RAB-mediated cultivation
- **Impact:** We have established a high-productivity, atmospheric CO₂ cultivation regime for *P. renovo*, enabling enhanced fuel yield and economics, and reduced carbon intensity.

Acknowledgements





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Ashton Zeller

UC San Diego

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Marcus Bray Lukas Dahlin Ryan Davis Jeff Linger Kim Rosenbach Matt Wiatrowski Mike Guarnieri

Additional Slides

Quad Chart Overview

UCSD has partnered with the Gross-Wen Technologies (GWT), Algix, and the National Renewable Energy Laboratory (NREL) to establish carbon sequestering molecular films for enhanced atmospheric CO_2 capture and increased biomass productivity. The project entitled "Biomolecular Films for Direct Air Captur of CO₂" will integrate core competencies from a partners, including computational modeling, protein and algal strain engineering, mass cultivation, biomas upgrading, and TEA/LCA in order to develop conversion process demonstrating the production c fuel intermediates and bioplastics from atmospheri CO_2 direct air capture (DAC). Successfi implementation of the proposed project offers dramati economic and sustainability benefits relative to conventional DAC systems.

Enabling bio-based DAC via molecular film technology

Key Personnel

UCSD: Karsten Zengler (PI) GWT: Martin Gross; Algix: Ashton Zeller; NREL: Mike Guarnieri

Program Summary

Period of performance:	Federal funds:	\$ 2M
36 months	Cost-share:	\$ 0.5M
	Total budget:	\$ 2.5M

	Key Milestones & Deliverables
Y1	• Establish baseline for <i>P. renovo</i> cultivation in GWT RAB system grown on atmospheric CO ₂ .
Y2	 Establish baseline for molecular film-mediated enhancement of atmospheric CO₂ capture in open pond systems. Achieve a 10% increase in algal biomass revenue potential, algal biomass quality <15% out of specification for downstream testing, 5% increase in productivity, 10% increase in CO₂ obtained from ambient air, and a 5% decrease in CO₂ costs via bio-based DAC.
¥3	 Achieve 25% increase in algal biomass revenue potential, algal biomass quality <10% out of specification for downstream conversion to fuel intermediates and bioplastics, a 10% increase in productivity, a 20% increase in CO₂ obtained from air, and a 10% decrease in CO₂ costs via bio-DAC Demonstrate upgrading of algal protein to bioplastics
• O at to su	Technology Impact ur bio-based, photoproduction approach offers a number of advantages over biotic DAC technologies; notably, little-to-no CAPEX/OPEX expenditures relative current and emerging abiotic CO ₂ capture and delivery systems, which require ubstantial water and energy inputs.
• O sy te	ur technology can be seamlessly integrated into conventional algal growth /stems that utilize either point source CO ₂ streams, or conventional DAC chnologies, synergistically reducing costs.

Responses to Previous Reviewers' Comments

N/A – Project was not previously reviewed.

Key Patents, Publications, and Presentations

Publications

 Dahlin LR, Guarnieri MT. (2021) Development of the high-productivity marine microalga, *Picochlorum renovo*, as a photosynthetic protein secretion platform. *Algal Research* 54, 102197

Patent Applications

• Photosynthetic protein secretion platform. US Patent App. 17/666,345

Presentations

- Dahlin LR. International Conference on Algal Biomass Biofuels, & Bioproducts. Waikoloa, HI, 2023.
- Guarnieri MT, et al. SIMB SBFC, Portland, OR, 2023.
- Dahlin LR. Algal Biomass Summit. Virtual. Oral Presentation. October 13th 2021.
- Dahlin L.R and Guarnieri, M.T. International Conference on Algal Biomass, Biofuels, & Bioproducts. Virtual. June 14-16 2021.
- Kim, Euihyun. Et al. Reconstruction of the Genome-scale Metabolic Model of *Picochlorum renovo*. GEAR: Guided Engineering Apprenticeship in Research.

References and Resources

- Algae farm design report: <u>https://www.nrel.gov/docs/fy16osti/64772.pdf</u>
- NREL algae farm TEA Excel tool: <u>https://www.nrel.gov/extranet/biorefinery/aspen-models/</u> (first set of files)
- NREL 2014 CAP design report: <u>https://www.nrel.gov/docs/fy14osti/62368.pdf</u>
- NREL 2011 biochemical ethanol design report: https://www.nrel.gov/docs/fy11osti/47764.pdf
- NREL 2015 SOT milestone: R. Davis, J. Markham, C. Kinchin, E. Tan, "2015 State of Technology Update." Internal BETO milestone report, Sept 30 2015 (rev3 re-issued Nov 24 2015)
- Dong et al. CAP publication (public reference that reflects key SOT parameters for CAP): T. Dong et al., "Combined algae processing: A novel integrated biorefinery process to produce algal biofuels and bioproducts." *Algal Research* 19 (2016): 316-323.
- Beal 2015: C. M. Beal et al., "Algal biofuel production for fuels and feed in a 100-ha facility: A comprehensive technoeconomic analysis and life cycle assessment." *Algal Research* 10 (2015): 266-279.
- Huntley 2015: M. E. Huntley et al., "Demonstrated large-scale production of marine microalgae for fuels and feed." *Algal Research* 10 (2015): 249-265.
- Lundquist 2010: T. J. Lundquist et al., "A realistic technology and engineering assessment of algae biofuel production." (2010); <u>http://www.energybiosciencesinstitute.org/media/AlgaeReportFINAL.pdf</u>
- NETL CO₂ carbon capture cost goals: <u>https://energy.gov/fe/science-innovation/carbon-capture-and-storage-research/carbon-capture-rd</u>
- USDA fertilizer pricing data: <u>https://quickstats.nass.usda.gov/</u>