

**2013 DOE Bioenergy Technologies** Office (BETO) Project Peer Review

**Pretreatment and Enzyme Hydrolysis** 



Tuesday, May 21<sup>st</sup>, 2013

**Technology Area Review: Biochemical Conversion** 

Principal Investigator: David K. Johnson

**Organization: National Renewable Energy Laboratory** 

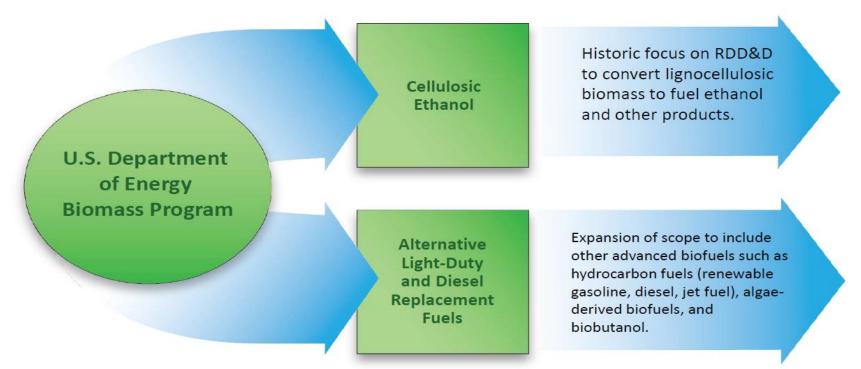
This presentation does not contain any proprietary, confidential, or otherwise restricted information

NREL is a national laboratory of the U.S. Department of Energy, Office of Energy Efficiency and Renewable Energy, operated by the Alliance for Sustainable Energy, LLC.

## **Project Overview**

Develop and apply fundamental knowledge of pretreatment and enzymatic hydrolysis concepts in process-relevant contexts to produce lignocellulosic biofuels

## **Expanding Scope**



## **Goal Statement**

To support the near-term Biochemical Platform cost and process performance goals:

•Generate cost-competitive <u>useable sugars</u> for conversion to cellulosic ethanol by 2012 (corn stover)

- Develop the <u>pretreatment/hydrolysis unit-operations</u> in a process relevant scaleable manner and so that they integrate well with downstream processes
- Reduce estimated mature technology processing costs for converting cellulosic feedstocks to ethanol

## Directly support BETO's Multi-Year Program Plan (MYPP) objectives (targets beyond 2012) :

• Produce sugars and other reactive intermediates "to support the 2017 goals for renewable gasoline, diesel, and jet fuel"

## **Quad Chart Overview**

## Timeline

- Project start date 2004
- Project end date 2017
- Percent complete 60%

## **Budget**

- Funding for FY11(\$6.5M / \$0)
- Funding for FY12(\$5.06M/\$0)
- Funding for FY13 (\$5.75M / \$0)
- Years the project has been funded – 10 years/ \$5.1M average annual funding.

## **Barriers**

- Bt-D Pretreatment Processing
- Bt-E Pretreatment Costs
- Bt-G Cellulase Enzyme Loading

## Partners

- Subcontracts
  - Auburn University, U. Georgia CCRC, Oklahoma State University, Baylor University, Virginia Tech University, U. Colorado, U. North Dakota, Colorado School of Mines, NIST, Washington State University
- Other Collaborations
  - North Carolina State University, Pennsylvania State University, U. Toronto, Andritz, IdeaCHEM,
- The project is managed under the Biochemical Platform at NREL

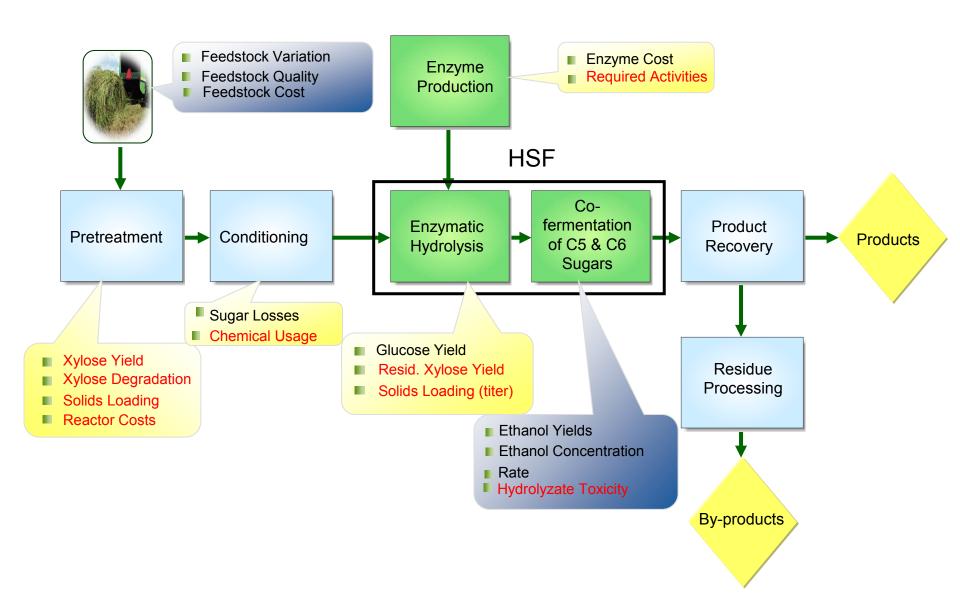
## Approach

### • Transition from knowledge development to unit-operation applications

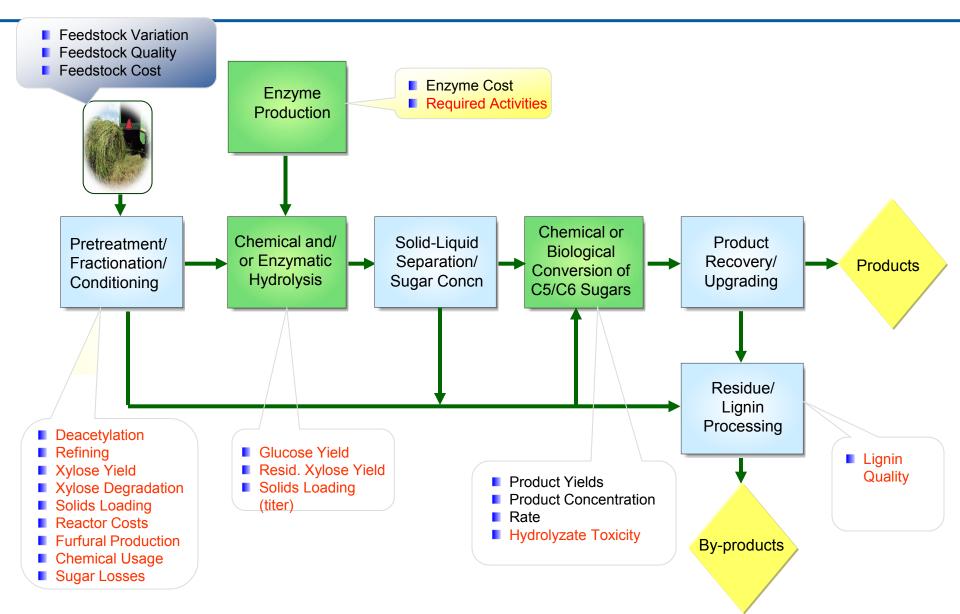
Knowledge Development	Applications
<ul> <li>Impact of pretreatment parameters on</li></ul>	<ul> <li>Selection of pretreatment configuration for 2012</li></ul>
monomeric xylose yields, inhibitor formation,	integrated biochemical conversion of corn stover to
and overall fermentation yields from high	ethanol, including TEA considerations to achieve
solids, unwashed slurries	2012 ethanol cost targets.
<ul> <li>Identification of approaches for conversion</li></ul>	Use of selected component enzymes to achieve
xylooligomers / residual xylan to monomer	xylooligomer saccharification
Rheological tools and methods for	<ul> <li>Application of rheological findings for bench-scale /</li></ul>
enzymatic hydrolysis slurries	pilot-scale high-solids saccharification reactor design
Identification of inhibitors and characterize     effects on biological and chemical processes	Mitigate toxicity of high-concentration hydrolyzates
<ul> <li>Investigate deoxygenation routes</li> <li>Investigate pathways to increase carbon</li></ul>	<ul> <li>Develop strategies for converting sugars into</li></ul>
chain length	hydrocarbon fuels and intermediates

- Management plan has well-defined performance targets leading to 2012 and 2017/2022 integrated process demonstrations
  - Detailed Annual Operating Plan (AOP) at Subtask Level
  - 10-12 milestones/deliverables per year
  - Tracking of results in annual State of Technology (SOT) updates

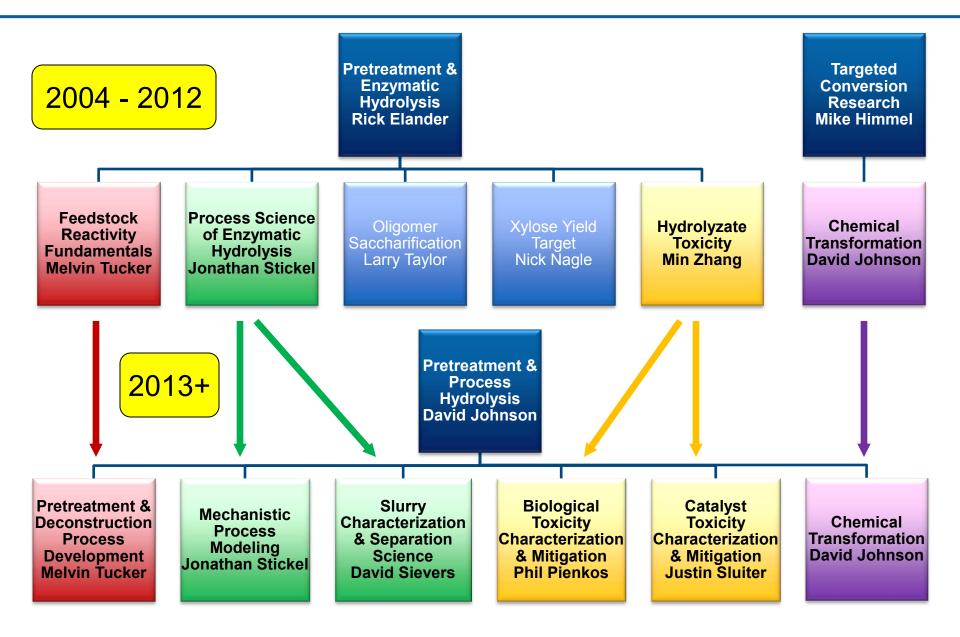
### **Biochemical Platform Process Impacts – Cellulosic Ethanol**



## **Biochemical Platform Process Impacts – Advanced Biofuels**



## **Task Structure**

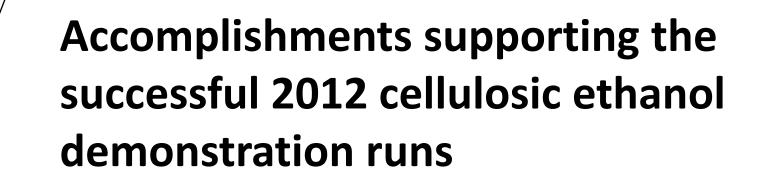




## Technical Accomplishments/Progress /Results

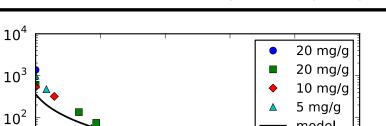
- Accomplishments supporting the successful 2012 cellulosic ethanol demonstration runs
- Progress aimed at further reducing the cost of converting biomass to sugars
- Progress in support of producing hydrocarbons from biomassderived sugars





## Characterization of Process Slurries

- Pretreated slurry rheology characterized
- Enzymatic hydrolysis rheological behavior characterized over reaction time
- Data used to specify enzymatic hydrolysis reactor vessels/agitators and pumps/pipelines in NREL's pilot plant
- Equipment successfully delivered performance to meet 2012 demonstration run goals



vield stress (Pa)

 $10^{1}$ 

10<sup>0</sup>

 $10^{-1}$ 



model

#### Process Science of Enzymatic Hydrolysis

 Deacetylated corn stover pretreated 150°C, 0.5% acid, as digestible as control CS pretreated 170°C, 1.0% acid, but furfural yield only 2% vs 10%

Deacetylation enabled use of lower severity pretreatment conditions with

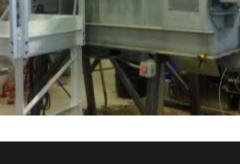
Pre-process with 0.1M NaOH, 80°C to remove acetate prior to pretreatment

• Equipment incorporating deacetylation and acid impregnation was scaled-up in support 2012 cellulose ethanol demonstration runs

2000 L Deacetylation/Impregnation Mix Tank



Continuous Dewatering Screw Press



## Deacetylation and Lower Severity Pretreatment of Corn Stover





# Identifying barriers to maximizing xylose yields

•In FY11 a series of xylooligomers with 4-Omethyl-α-glucuronic acid residue (MeGlcA) identified as most abundant pool of remaining convertible xylose

•X-MeGlcA and X-X-MeGlcA predominant species. Persist despite thermochemical oligomer hold, or enzymatic hydrolysis

•FY12 D milestone: achieve 50% conversion of 4-O-methyl-α-glucuronic acid-substituted xylooligomers to monomeric xylose from relevant pretreatment liquid

•Of four tested enzymes GH67s from *A. niger* and *Geobacillus stearothermophilus* exceeded target reductions of X-MeGIcA and X-X-MeGIcA

#### Composition of Deacetylated Hydrolyzate

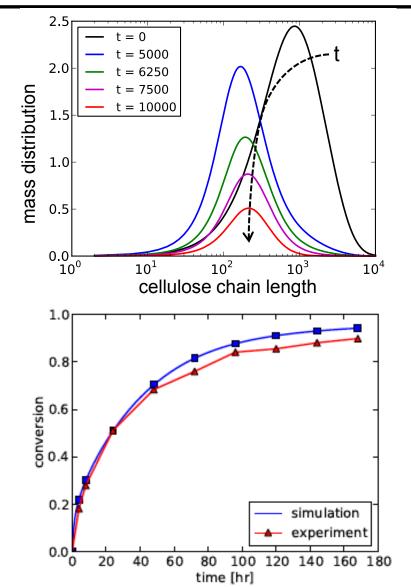
	Concentration	
	(mg/mL)	
Glucose	1.8	
Celloligomers	3.8	
Xylose	0.8	
Xylooligomers	15.2	

## Conversion of Xylooligomers using Alpha-Glucuronidases

$\alpha$ -glucuronidase	X-X-MeGlcA	X-MeGlcA
GH115 <sub>B</sub>	3	18
GH67 <sub>c</sub>	0	15
GH67 <sub>G</sub>	77	87
GH67 <sub>A</sub>	96	83
No $\alpha$ -glucuronidase	2	6
no enzyme	0	0

## Modeling of Enzymatic Hydrolysis Kinetics

- Models incorporating the independent action of different enzyme types (EG, CBH, βG) used to predict the population distribution of cellulose
- Simulation closely predicts actual hydrolysis conversion over time
- Useful for predicting conversion yields in process simulation based on enzyme cocktail and feedstock properties



FY11 D-Milestone, "Incorporate additional phenomena in the enzymatic hydrolysis kinetics model and perform model validation with experimental data"

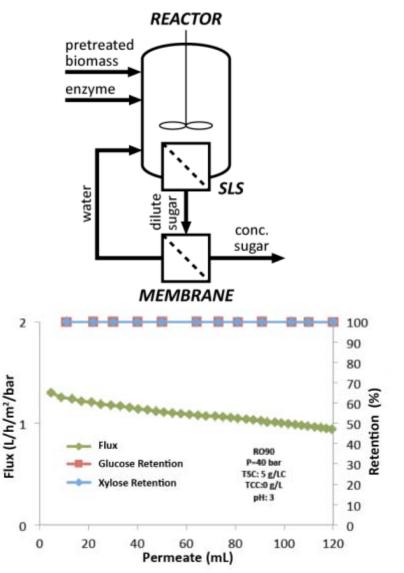
NATIONAL RENEWABLE ENERGY LABORATORY

#### Process Science of Enzymatic Hydrolysis

## Separations Development for Continuous Enzymatic Hydrolysis

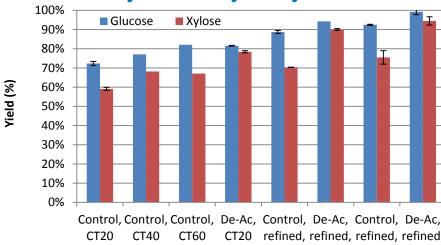
#### Slurry Characterization and Separation Science

- Solid-liquid separation (SLS) to remove products: reduce product inhibition and increase productivity
  - Development and experimentation planned for FY13 through FY15
- Sugar product concentrated to prepare sugar stream for variety of fuel pathways.
  - High sugar retention demonstrated with nanofiltration membrane



FY13 D-Milestone, "Develop membrane-based water removal (sugar concentrating) process technology."

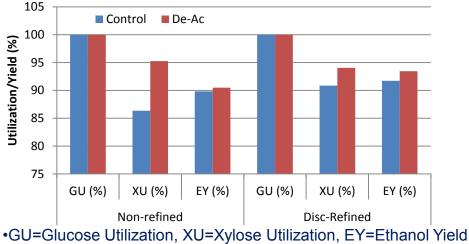
### Deacetylation and Mechanical Refining with Dilute-Acid Pretreatment



#### Enzymatic hydrolysis

CT20 refined, refined, refined, refined, CT10 CT10 CT20 CT20

#### **Fermentation**



- Pre-process corn stover with 0.1M NaOH at 80 °C for 2h to remove acetate prior to pretreatment
- Post-pretreatment mechanical • refining requires less energy, and increases cellulose accessibility
- Deacetylation and mechanical refining significantly enhance enzymatic digestibility of low severity dilute-acid pretreated corn stover (150 °C, 0.5% acid, 20 min)
- Deacetylation significantly increases xylose utilization in fermentation and improves ethanol yield and titer
- Deacetylation and mechanical • refining can reduce the minimum ethanol selling price

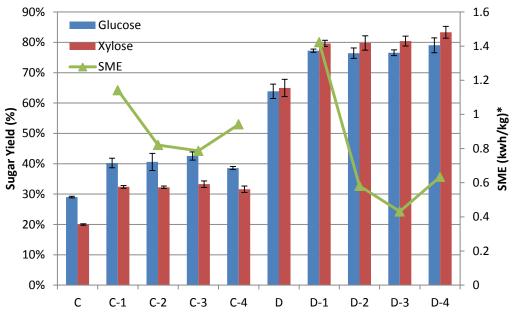
# Deacetylation and Mechanical Refining without Dilute Acid Pretreatment

#### Pretreatment & Deconstruction Process Development





A Coperion ZSK-25 twin screw, co-rotating extruder from IdeaCHEM, Rapid City, SD



- C-control, non-extruded; C1-C4- control, extruded
- D-deacetylated, non-extruded; D1-D4-deacetylated, extruded
- Total Enzyme loading=26mg/g cellulose.
- SME = Specific Mechanical Energy (kWh/kg)

Mechanical refining with deacetylation could potentially replace acid pretreatment to achieve high sugar yields and produce a clean sugar stream without degradation products.

Similar results obtained with PFI mill and disk refiner

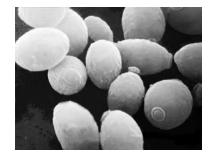




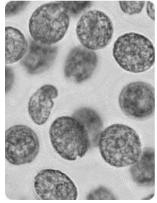
### Identification of Inhibitors to Fuel Producing Organisms

#### **Biological Toxicity Characterization & Mitigation**

- Identify specific inhibitors and assess impacts on HC producing microorganisms
- Ultimately develop strategy to mitigate their impact on the biological processes involved.
- Identify production pathways and relevant strains
  - Fatty Acid Pathway
  - Isoprenoid Pathway
  - Polyhydroxybutyrate Pathway
  - Polyketide Pathway
- Determine growth characteristics using acetate w/ or w/o sugars
- Develop high throughput inhibition assay
- Implement high throughput method to generate inhibitor profiles.







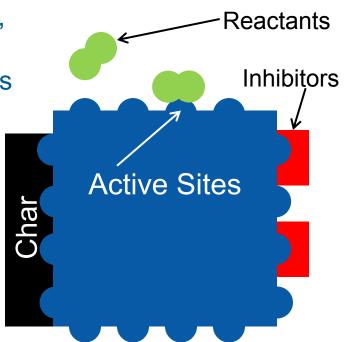


#### Prediction of Catalyst Deactivation

- Analysis for fermentation processes have been focused on carbohydrates and biological inhibitors.
- Many potential catalytic inhibitors are not routinely or robustly measured by current methods.
- Need to learn how biomass pre-processing, pretreatment, and hydrolysis should be optimized for catalytic conversion processes

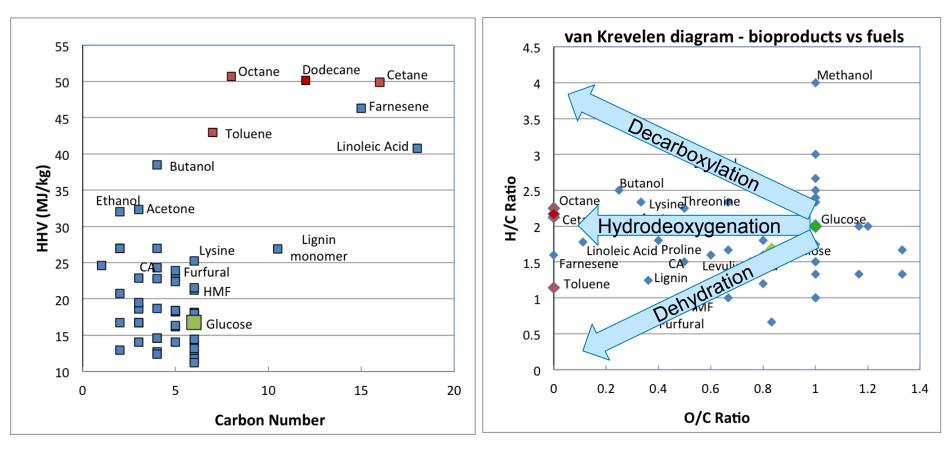
This task is developing and adapting methods for routine quantification of potential catalyst inhibitors.

- ICP for detection of known toxic elements (i.e. sulfur)
- LC/MS & GC/MS for quantification of potential char producing, non-carbohydrate components (i.e. phenolic or aldehydes)



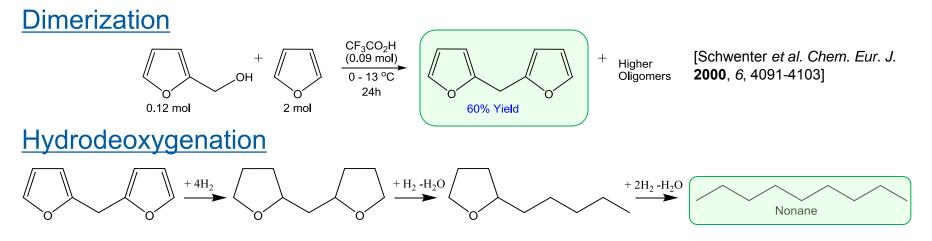
### Production of Hydrocarbon Fuels from Biomass-derived Components

- Increase energy content Decrease oxygen content
- Increase carbon chain length favors jet and diesel over gasoline



Top Value Added Chemicals from Biomass Volume I - Results of Screening for Potential Candidates from Sugars and Synthesis Gas (2004). Top 30 chemicals + others

## Furans to Hydrocarbons



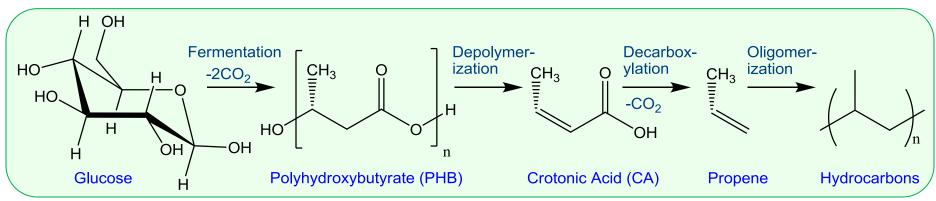
- Can a furfural-based dimer be produced and hydrodeoxygenated to alkanes
- Moderate yields of C7 C9 alkanes were made from furfuryl alcohol furan dimer
- With much higher yields it could be possible to produce HC fuels from furfural

Catalyst	T (°C)	t (h)	Oxygenates %	Heptane %	Octane %	Nonane %
Pt/C	165-300	1	60	5	6	5
	300	4	54	12	23	11
Pt/C Pt/Al <sub>2</sub> O <sub>3</sub>	300	3	76	1	4	2
Pt/Al <sub>2</sub> O <sub>3</sub>	300-350	5	37	13	28	11

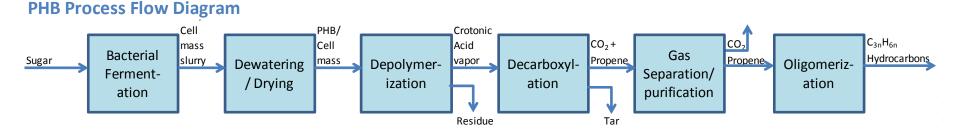
2012 D-Milestone. Evaluate experimentally the transformation of furfuraldehyde into a mixture of aliphatic hydrocarbons

## Polyhydroxyalkanoates to Hydrocarbons

#### **Chemical Transformation**



- Fermentation gives solid product containing up to 80% PHB
- Depolymerization and decarboxylation can be combined to give propene a gaseous product in 70-80% molar yield
- Known technology would be used to separate propene and CO<sub>2</sub>
- Propene oligomerization using known technology to give HC fuels



## Polyhydroxyalkanoates to Hydrocarbons

- Can PHB be converted to propene in a simple reaction system
- A simple heated system without catalyst or solvent was shown to convert the intermediate CA to propene and CO<sub>2</sub>
- Commercial PHB and PHB in bacteria cells also converted to propene
- Less costly process without prior PHB isolation appears possible

Substrate	Temperature (°C)	Pressure (psig) *	Propene Yield (molar %)	CO <sub>2</sub> Yield (molar %)
СА	350	$23 \pm 3$	$19 \pm 3$	$15 \pm 7$
СА	400	76 ± 2	$77 \pm 3$	74 ± 3
PHB	350	$24 \pm 1$	$22 \pm 3$	$20 \pm 2$
РНВ	400	71 ± 7	70 ± 7	$78 \pm 6$
C. Necator 75% PHB	400	28	69	118

Reaction Time 15 min; At least 5 replicate reactions except for C. Necator

\* Autogenic pressure at room temperature

2012 D-Milestone. Identify process conditions to achieve simultaneous depolymerization and thermocatalytic conversion of a PHA into a hydrocarbon)

NATIONAL RENEWABLE ENERGY LABORATORY

## Relevance

- Task goals have been and will continue to be clearly tied to wellestablished Biochemical Platform technical and economic goals
  - Sugar yields from pretreatment and enzymatic hydrolysis unit operations
  - Relevant processes, equipment, and mode of operation taken to larger scale
  - Considers impacts on downstream operations sugar concentration, inhibitor formation, lignin quality
- Findings and know-how were directly utilized in 2012 biochemical State-of-Technology process demonstrations
- Transitioning to MYPP goal of by 2022, achieving the overall Program performance goal of \$3 per GGE (\$2011), based on data at the integrated pilot scale
- By 2017, validate integrated production of a hydrocarbon fuel or blend stock from cellulosic biomass via a biological or chemical route at integrated bench-scale
- Develop commercially viable technologies for reducing the processing cost of converting biomass feedstocks into energy dense fungible, liquid transportation fuels, bioproducts, and chemical intermediates

## **Critical Success Factors**

Factors	Strategy to Overcome
Technical	<ul> <li>Develop and apply knowledge on how to improve pretreatment AND enzymatic hydrolysis as a SYSTEM to saccharify biomass         <ul> <li>Coordinated development to achieve BETO conversion targets</li> <li>Well-defined technical targets and schedule to achieve overall targets</li> </ul> </li> </ul>
Business/ Market	<ul> <li>Numerous collaborative projects and proposals are leveraging core programmatic pretreatment/enzymatic hydrolysis capabilities and expertise</li> <li>Prototype process equipment design and testing (vendor market)</li> <li>Publications/outreach to disseminate R&amp;D findings</li> </ul>
Regulatory	<ul> <li>High solids loadings and concentrations to reduce water usage</li> <li>Provide data and samples to evaluate wastewater treatment and solids disposal</li> </ul>







## Challenges

- Development of scaleable unit operations producing soluble sugar/carbon streams for cost-effective biological/chemical production of hydrocarbons from biomass
  - Will require further reduction in cost of producing sugars
  - Further reduction in formation of degradation products in pilot-scale, continuous pretreatment systems
  - Integration of sugar production with organisms and catalysts for producing new fuels possibly involving new separations and concentrating processes
  - Pretreatment must be coordinated with production of lignin-derived products
- Achieving process performance targets for new fuels at process relevant scale probably with new reactor systems







## Future Work – Reduced Sugar Costs

- Pretreatment and Deconstruction Process
   Development
  - Optimize / scale-up mechanical refining of deacetylated feedstock from low severity or no acid pretreatments
  - Optimize furfural production and recovery in reactor flash or separate two stage reactor processes
- Slurry Characterization & Separation Science



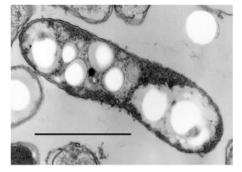


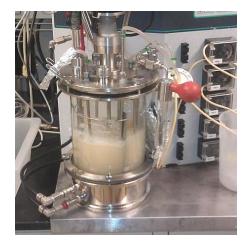
- Further develop separation methods to enable continuous enzymatic hydrolysis
- Develop methods for characterizing wet-granular slurries and correlate with pretreatment performance
- Mechanistic Process Modeling
  - Parameter estimation and experimental validation of model describing cellulose enzymatic hydrolysis kinetics
  - Continue development of CFD for mixing biomass slurries and couple with reaction kinetics models

## Future Work – Hydrocarbon Fuels

### Biological Toxicity Characterization & Mitigation

- Perform inorganic and detailed organic analysis of saccharified slurries to identify potential inhibitors and relevant concentration ranges.
- Identify top three inhibitors for select microorganisms.
- <u>Catalyst Toxicity Characterization & Mitigation</u>
  - Develop small scale catalyst bed for testing individual analytes to determine magnitudes of catalyst poisoning
  - Evaluate methods for removal or mitigation of the inhibitors in hydrolyzates prior to catalytic upgrading
- <u>Chemical Transformation</u>
  - Further develop processes for oligomerization of furfural derived chemicals and their hydrodeoxygenation to hydrocarbons
  - Scale-up conversion of PHB to propene and demonstrate production of PHB on biomass hydrolyzates
  - Perform TEA of furfural and PHB processes





## Summary

- Work in the Pretreatment and Enzymatic Hydrolysis Task was critical to achieving 2012 technical performance and ethanol cost targets
  - Identification of pretreatment configuration for 2012 integrated biochemical conversion of corn stover to ethanol
  - Scale-up of deacetylation and its impact on decreased severity pretreatments with lower acid loadings, decreased neutralization chemicals, and improved enzyme digestibilities
  - Identification and mitigation of the fermentation inhibitors formed in pretreatment

- Work in Task is transitioning to achieving the 2022 goal of \$3 per GGE (\$2011), for hydrocarbon fuel or fuel blend stock from cellulosic biomass
  - Modifications to pretreatment aimed at lowering sugar production costs
  - Evaluation of hydrolyzate toxicity towards new hydrocarbon and intermediate producing organisms (and in the near future catalysts)
  - Evaluation of processes for converting sugars and furfurals into hydrocarbons

## **Acknowledgements**

- Subtask Leaders
  - Steve Decker
  - Nick Nagle
  - Philip Pienkos
  - Jonathan Stickel
     J. Farmer
  - David Sievers
  - Justin Sluiter
  - Melvin Tucker
  - Min Zhang
- Staff
  - B. Adney
  - G. Beckham
  - S. Black
  - B. Bray

- Staff (continued)
  - R. Brunecky
  - X. Chen
  - C. Dibble
- M. Franden
  - A. Griggs
  - S. Habas
  - W. Hjelm
  - D Humbird
  - T. Johnston
  - E. Kuhn
  - J. Lischeske
  - W. Michener

- Staff (continued)
  - A Mittal
  - L. Moens
  - A. Mohagheghi
  - H. Pilath
  - M. Resch
  - J. Shekiro
  - D. Sievers
  - D. Templeton
  - T Vinzant
  - W. Wang
  - J. Wolfe



## Funding

US DOE EERE BioEnergy Technology Office

## **Additional Slides**

### **Previous Reviewers Comments**

## 2011 Biochemical Platform Peer Review

#### Comment

#### Response

Comprehension of all parts of the project at once is difficult. Periodic economic evaluations of contemplated process additions might provide a framework for this.

The milestones are well defined however no risk mitigation was addressed.

We attempt to include understanding of the process implications of pretreatment approaches and conditions across relevant downstream conversion elements. This clearly includes the impact on enzymatic hydrolysis, including digestibility of cellulose and any remaining insoluble xylan and/oligomeric soluble xylose after pretreatment. Not so directly, it also includes understanding the impact of pretreatment upon fermentation of released sugars, especially as related to the presence of compounds solubilized from biomass during pretreatment and enzymatic hydrolysis, as well as generated via undesirable sugar degradation reactions during pretreatment. All of these factors are well understood in terms of their impact on overall process economics, which is routinely evaluated via annual State of Technology updates as well as by specific technoeconomic impact studies that are incorporated into many task milestones. These process economic evaluations are a key risk mitigation strategy to focus efforts on the most economically important factors.

This research has a long history in trying to understand dilute acidenzyme hydrolysis kinetics and increase in sugar yields. It seems that NREL has for years been doing the same thing. That being said, however, it's clear that the difference in the latest research is that they are doing the work at larger scale which brings new equipment changes and different technical challenges. In regard to the continued use of dilute-acid pretreatment as the "standard" pretreatment method and the incremental progress related to that technology. We agree that our R&D plan is focused on incremental improvements, especially for xylose yields, as much is already well understood about dilute acid pretreatment mechanisms and reaction kinetics. The reviewers correctly pointed out the challenge of translating these improvements to pilot-scale, continuous systems and the drive toward lower acid usage levels by the use of Fe salts and other approaches, which can help achieve these incremental improvements at reaction conditions that are better suited towards conditions that are more practical for implementation at this scale of operation.

### **Previous Reviewers Comments**

## 2011 Biochemical Platform Peer Review

#### Comment

#### Response

Success will ultimately depend on the translation of this research in a scaled up demonstration and application. Consideration of water treatment and recycling could add to the challenges of process integration.

Reviewers noted that the application of much of the presented work in scale-up applications was a critical success factor, with a specific mention of water recycling/treatment. Such factors are specifically being addressed within the Biochemical Processing Integration Task, which was presented later in the review meeting. We strive for well-integrated activities across this task and the Biochemical Processing Integration Task, as that becomes the ultimate proving ground of process concepts in a truly integrated process.

## Publications and Presentations: FY2011-13

#### **Publications**

Kamireddy, S. R.; Li, J.; Tucker, M.; Degenstein, J.; Ji, <u>(2013). Effects and Mechanism of Metal Chloride Salts on Pretreatment</u> and Enzymatic Digestibility of Corn Stover. Industrial and Engineering Chemistry Research. Vol. 52(5), 6 February 2013; pp. 1775-1782; NREL Report No. JA-5100-58138. <u>http://dx.doi.org/10.1021/ie3019609</u>

Huang XF, Santhanam N, Badri DV, Hunter WJ, Manter DK, Decker SR, Vivanco JM, Reardon KF. (2013). "Isolation and characterization of lignin-degrading bacteria from rainforest soils." Biotechnol Bioeng. Jan 7. doi: 10.1002/bit.24833. [Epub ahead of print]

Wei, H.; Tucker, M. P.; Baker, J. O.; Harris, M.; Luo, Y. H.; Xu, Q.; Himmel, M. E.; Ding, S. Y. <u>(2012). Tracking Dynamics of</u> <u>Plant Biomass Composting by Changes in Substrate Structure, Microbial Community, and Enzyme Activity.</u> Biotechnology for Biofuels. Vol. 5(4), April 2012; 20 pp.; NREL Report No. JA-2700-56077. <u>http://dx.doi.org/10.1186/1754-6834-5-20</u>

Chen, X.; Tao, L.; Shekiro, J.; Mohagheghi, A.; Decker, S.; Wang, W.; Smith, H.; Park, S.; Himmel, M. E.; Tucker, M. (2012). Improved Ethanol Yield and Reduced Minimum Ethanol Selling Price (MESP) by Modifying Low Severity Dilute Acid Pretreatment with Deacetylation and Mechanical Refining: 1) Experimental. Biotechnology for Biofuels. Vol. 5, August 2012; 10 pp.; NREL Report No. JA-5100-55366. http://dx.doi.org/10.1186/1754-6834-5-60

Selig, M., C-W. Hsieh, C. Felby, L. Thygesen, G. Turner, M. E. Himmel, S. R. Decker. (2012). "Considering water activity and the effect of soluble species on high solids saccharification of lignocellulosic biomass". Biotechnol Prog. 28(6):1478-90.

Santhanam, N., D. V. Badri, S. R. Decker, D. K. Manter, K. F. Reardon, and J. M. Vivanco. (2012). "Lignocellulose decomposition by microbial secretions". In: F. Baluška and J. Vivanco (eds). Secretions and Exudates in Biological Systems. Springer, New York. pp.125-153.

Shekiro, J.; Kuhn, E. M.; Selig, M. J.; Nagle, N. J.; Decker, S. R.; Elander, R. T. (2012). Enzymatic Conversion of Xylan Residues from Dilute Acid-Pretreated Corn Stover. Applied Biochemistry and Biotechnology. Vol. 168(2), September 2012; pp. 421-433; NREL Report No. JA-5100-52347. <u>http://dx.doi.org/10.1007/s12010-012-9786-5</u>

Tao, L.; Chen, X.; Aden, A.; Kuhn, E.; Himmel, M. E.; Tucker, M.; Franden, M. A. A.; Zhang, M.; Johnson, D. K.; Dowe, N.; Elander, R. T. (2012). Improved Ethanol Yield and Reduced Minimum Ethanol Selling Price (MESP) by Modifying Low Severity Dilute Acid Pretreatment with Deacetylation and Mechanical Refining: 2) Techno-Economic Analysis. Biotechnology for Biofuels. Vol. 5, September 2012; 11 pp.; NREL Report No. JA-5100-55704. <u>http://dx.doi.org/10.1186/1754-6834-5-69</u>

## Publications and Presentations: FY2011-13

Publications (continued)

Chen, X.; Shekiro, J.; Elander, R.; Tucker, M. (2012). Improved Xylan Hydrolysis of Corn Stover by Deacetylation with High Solids Dilute Acid Pretreatment. Industrial and Engineering Chemistry Research. Vol. 51(1), 11 January 2012; pp. 70-76; NREL Report No. JA-5100-50960. <u>http://dx.doi.org/10.1021/ie201493g</u>

Griggs, A. J.; Stickel, J. J.; Lischeske, J. J. (2012). Mechanistic Model for Enzymatic Saccharification of Cellulose using Continuous Distribution Kinetics II: Cooperative Enzyme Action, Solution Kinetics, and Product Inhibition. Biotechnology and Bioengineering. Vol. 109(3), March 2012; pp. 676-685; NREL Report No. JA-5100-51652. http://dx.doi.org/10.1002/bit.23354

Griggs, A. J.; Stickel, J. J.; Lischeske, J. J. (2012). Mechanistic Model for Enzymatic Saccharification of Cellulose using Continuous Distribution Kinetics I: Depolymerization by EGI and CBHI. Biotechnology and Bioengineering. Vol. 109(3), March 2012; pp. 665-675; NREL Report No. JA-5100-51651. <u>http://dx.doi.org/10.1002/bit.23355</u>

Stickel, J. J.; Griggs, A. J. (2012). Mathematical Modeling of Chain-End Scission using Continuous Distribution Kinetics. Chemical Engineering Science. Vol. 68(1), January 2012; pp. 656-659; NREL Report No. JA-5100-49632. http://dx.doi.org/10.1016/j.ces.2011.09.028

Santhanam, N., D. V. Badri, S. R. Decker, D. K. Manter, K. F. Reardon, and J. M. Vivanco. (2012). "Lignocellulose decomposition by microbial secretions". In: F. Baluška and J. Vivanco (eds). Secretions and Exudates in Biological Systems. Springer, New York. pp.125-153.

Segato, F., A. Damasio, T. Gonçalves, R. de Lucas, F. Squina, S. R. Decker. (2012). "Secretion of client proteins in Aspergillus". Enzyme and Microbial Technology. 51:100-106

Goldberg, R. N., B. E. Lang, B. Coxon, and S. R. Decker. (2011). "Saturation molalities and standard molar enthalpies of solution of  $\alpha$ -D-xylose(cr) in H2O(I); Standard molar enthalpies of solution of 1,4- $\beta$ -D-xylobiose(am), and 1,4- $\beta$ -D-xylotriose(am) in H2O(I)". J. Chem. Thermodynam. 29:480-489

Santhanam, N., J. M. Vivanco, S. R. Decker, and K. F. Reardon. (2011-online). "Engineered expression of industrially relevant laccases -- prokaryotic style". Trends Biotechnol. 29:480-489.

Tao, L., A. Aden, R. T. Elander, V. R. Pallapolu, Y.Y. Lee, R. J. Garlock, V. Balan, B. E. Dale, Y. Kim, N. S. Mosier, M. R. Ladisch, M. Falls, M. T. Holtzapple, R. Sierra, J. Shi, M. A. Ebrik, T. Redmond, B. Yang, C. E. Wyman, B. Hames, *et al.* (2011). "Process and technoeconomic analysis of leading pretreatment technologies for lignocellulosic ethanol production using switchgrass". *Bioresource Technology*. doi:10.1016/j.biortech.2011.07.051

Zhu, Y. M.; Malten, M.; Torry-Smith, M.; McMillan, J. D.; Stickel, J. J. (2011). Calculating Sugar Yields in High Solids Hydrolysis of Biomass. Bioresource Technology. Vol. 102(3), February 2011; pp. 2897-2903; NREL Report No. JA-5100-49739. http://dx.doi.org/10.1016/j.biortech.2010.10.134

## Publications and Presentations: FY2011-13

#### Publications (continued)

Dibble, C. J.; Shatova, T. A.; Jorgenson, J. L.; Stickel, J. J. (2011). Particle Morphology Characterization and Manipulation in <u>Biomass Slurries and the Effect on Rheological Properties and Enzymatic Conversion</u>. Biotechnology Progress. Vol. 27(6), November/December 2011; pp. 1751-1759; NREL Report No. JA-5100-51065. <u>http://dx.doi.org/10.1002/btpr.669</u>

Chen, X.; Shekiro, J.; Franden, M. A.; Wang, W.; Johnson, D. K.; Zhang, M.; Kuhn, E.; Tucker, M. P. (2011). Impacts of <u>Deacetylation Prior to Dilute Acid Pretreatment on the Bioethanol Process</u>. Biotechnology for Biofuels. Vol. 5(1), December 2011; 8 pp.; NREL Report No. JA-5100-53651. <u>http://dx.doi.org/10.1186/1754-6834-5-8</u>

Selig, M. J.; Tucker, M. P.; Law, C.; Doeppke, C.; Himmel, M. E.; Decker, S. R. <u>(2011). High-Throughput Determination of</u> <u>Glucan and Xylan Fractions in Lignocelluloses.</u> Biotechnology Letters. Vol. 33(5), May 2011; pp. 961-967; NREL Report No. JA-2700-49134. <u>http://dx.doi.org/10.1007/s10529-011-0526-7</u>

Wei, H.; Donohoe, B. S.; Vinzant, T. B.; Ciesielski, P. N.; Wang, W.; Gedvilas, L. M.; Zeng, Y.; Johnson, D. K.; Ding, S. Y.; Himmel, M. E.; Tucker, M. P. (2011). Elucidating the Role of Ferrous Ion Cocatalyst in Enhancing Dilute Acid Pretreatment of Lignocellulosic Biomass. Biotechnology for Biofuels. Vol. 4, 2011; 16 pp.; NREL Report No. JA-2700-52200. http://dx.doi.org/10.1186/1754-6834-4-48

Degenstein, J. C.; Kamireddy, S.; Tucker, M. P.; Ji, Yun (2011). Novel Batch Reactor for the Dilute Acid Pretreatment of <u>Lignocellulosic Feedstocks with Improved Heating and Cooling Kinetics</u>. International Journal of Chemical Reactor Engineering. Vol. 9, January 2011; pg. A95; NREL Report No. JA-5100-53358.

Donohoe, B. S.; Vinzant, T. B.; Elander, R. T.; Pallapolu, V. R.; Lee, Y. Y.; Garlock, R. J.; Balan, V.; Dale, B. E.; Kim, Y.; Mosier, N. S.; Ladisch, M. R.; Falls, M.; Holtzapple, M. T.; Sierra-Ramirez, R.; Shi, J.; Ebrik, M. A.; Redmond, T.; Yang, B.; Wyman, C. E.; Hames, B. (2011). Surface and Ultrastructural Characterization of Raw and Pretreated Switchgrass. Bioresource Technology. Vol. 102(24), December 2011; pp. 11097-11104; NREL Report No. JA-2700-53545. http://dx.doi.org/10.1016/j.biortech.2011.03.092

Tao, L.; Aden, A.; Elander, R. T.; Pallapolu, V. R.; Lee, Y. Y.; Garlock, R. J.; Balan, V.; Dale, B. E.; Kim, Y.; Mosier, N. S.; Ladisch, M. R.; Falls, M.; Holtzapple, M. T.; Sierra, R.; Shi, J.; Ebrik, M. A.; Redmond, T.; Yang, B.; Wyman, C. E.; Hames, B. (2011). Process and Technoeconomic Analysis of Leading Pretreatment Technologies for Lignocellulosic Ethanol Production Using Switchgrass. Bioresource Technology. Vol. 102(24), December 2011; pp. 11105-11114; NREL Report No. JA-5100-51238. http://dx.doi.org/10.1016/j.biortech.2011.07.051

## Publications and Presentations: FY2011-13

Publications (continued)

Sandoval, N. R.; Mills, T. Y.; Zhang, M.; Gill, R. T. (2011). Elucidating Acetate Tolerance in E. coli Using a Genome-Wide Approach. Metabolic Engineering. Vol. 13(2), March 2011; pp. 214-224; NREL Report No. JA-5100-51615. http://dx.doi.org/10.1016/j.ymben.2010.12.001

Helm, R. F.; Jervis, J.; Ray, W. K.; Willoughby, N.; Irvin, B.; Hastie, J.; Schell, D. J.; Nagle, N. <u>(2010). Mass Spectral Analyses</u> of Corn Stover Prehydrolysates to Assess Conditioning Processes. Journal of Agricultural and Food Chemistry. Vol. 58(24), 22 December 2010; pp. 12642-12649; NREL Report No. JA-5100-51595. <u>http://dx.doi.org/10.1021/jf1031197</u>

Zhu Y., Malten M., Torry-Smith M., McMillan J.D., Stickel J.J. (2010). Calculating sugar yields in high solids hydrolysis of biomass. Bioresour. Technol., 102:2897–2903.

Numerous oral and poster presentations have been made at ACS and AIChE Annual Meetings, annual Symposia on Biotechnology for Fuels and Chemicals, and annual meetings of the Society of Rheology.

# **Supplementary Slides**

## Milestone Schedule for Pretreatment and Enzyme Hydrolysis Task

#### FY 2012

Туре	Title	Due Date
E	Identify gene targets for improved resistance to acetate and furfural	10/31/11
D	Select overall pretreatment configuration for 2012 integrated	12/31/11
	biochemical conversion of corn stover to ethanol, including	
	technoeconomic considerations of any pre-processing and post-	
	processing steps being used to achieve 2012 pretreatment and	
	enzymatic hydrolysis yield targets.	
D	Implement coupled computational fluid dynamics and kinetics	03/31/12
	simulations for enzymatic hydrolysis at moderate solids loadings and	
	validate against appropriate experimental data.	
D	Parametric study using continuous, pilot-scale pretreatment reactor	05/31/12
	system on corn stover to determine operating conditions that achieve	
	overall process yields of 90% of monomeric xylose with ≤5% conversion	
	to furfural	
Е	Determine pore-size distribution of milled and pretreated biomass	06/30/12
	(likely corn stover and pine). Determine convective and diffusive	
	transport of solutes (salts, sugars, and macromolecules) in packed	
	beds of the biomass.	
Е	Produce at least 50 kg (dry) of pretreated un-deacetylated and	06/30/12
	deacetylated corn stover feedstocks in a continuous pretreatment	
	reactor for commercial scale disk refining. Determine pretreatment	
	and enzymatic digestibility yields, feedstock reactivities, and refining	
	energies at various refining conditions.	
Е	Identify limit products which act as barriers to achieving 90% glucan	07/15/12
	conversion and 80% xylan conversion in 15-30% total solids pretreated	
	corn stover digestions.	
Е	Knockout and overexpression of furfural toxicity related genes in	07/15/12
	Zymomonas mobilis strains and evaluate their impact on furfural	
	toxicity.	
D/E	Achieve 50% conversion of 4-O-methyl-alpha glucuronic acid-	08/31/12
	substituted xylooligomers to monomeric xylose from relevant	
	pretreatment liquid	
D/E	Evaluate impact of candidate genes related to acetate and furfural	09/30/12
	toxicity contribution to xylose fermentation of Zymomonas mobilis	
	strains	

#### FY 2013

-		1
Туре	Title	Due Date
D	Develop membrane-based water removal (sugar concentrating)	02/28/13
	process technology	
Е	Report kinetics of depolymerization and decarboxylation reactions	03/31/13
	involved in conversion of polyhydroxybutyrate to propene	
Е	Evaluate various mathematical modeling approaches for dilute-acid	03/31/13
	pretreatment of lignocellulosic biomass and select one that includes	
	the most relevant chemical and physical mechanisms	
Е	Assess the possibility of utilizing acetate in hydrolysate/slurry along	03/31/13
	with sugars or pre-hydrolysate liquor from biomass deacetylation with	
	hydrocarbon producing organisms	
Е	Demonstrate experimentally a new pathway from a furfuraldehyde to	06/30/13
	a liquid hydrocarbon with a chain length of C9 or higher	
Е	Develop experimental and data analysis methods for routinely	06/30/13
	measuring the rheological properties of biomass slurries by means of	
	large amplitude shear rheometry	
Е	Investigate various co-catalysts in hydrothermal, dilute acid, and	07/31/13
	alkali pretreatment for the production of furfural/HMF for upgrading to	
	C-10 to C-20 hydrocarbons	
D	Scale up promising biomass hydrolysis/deconstruction technology to a	08/31/13
	continuous reactor to produce intermediates for supply to the	
	Chemical Transformation subtask and other stakeholders for	
	conversion to hydrocarbon fuel molecules.	
D	Demonstrate production of a hydrocarbon fuel intermediate from a	09/30/13
	process relevant lignocellulosic derived sugar stream	
D	Utilize existing software and develop additional computational tools	09/30/13
	to enable full-scale simulation of a mixing and reacting enzymatic	
	hydrolysis reactor	
D	Identify inhibitors and their impact on model hydrocarbon fuel and/or	09/30/13
	intermediate-producing microorganisms in hydrolysate liquor and/or	
	saccharified slurry from dilute acid pretreatment	
D	Develop a method for quantification of organic inhibitors to catalytic	09/30/13
	conversion present in dilute acid pretreatment liquors	

### Lower cost of producing sugars

- Lower reactor costs
  - Decreased pretreatment severity and temperature
- Lower chemical usage and costs
  - Decreased sulfuric acid and NH<sub>4</sub>OH usage
- Decrease inhibitors in fermentation
  - Deacetylation decreased acetate in fermentation
  - Low severity pretreatments lowered salts, furfural, HMF concentration
- Mechanical refining improves enzymatic digestibility of low severity pretreatments
  - Bench scale: PFI mill, blender, disk refining, twin screw extruding
  - Industrial scale: Andritz 36-inch disk refiner, Szego mill
- Recover furfural and acetate in reactor flash for downstream conversion
- Developed new process of deacetylation of corn stover followed by mechanical refining for high enzymatic digestibility WITHOUT pretreatment
  - Sugar yields 80% although with slightly higher enzyme dosage

Develop processes for low cost reactive lignin

- Low severity pretreatment lessen lignin condensation
- Deacetylation provides source of acetate and soluble lignin

## **Support of FY12 SOT Demonstration**

### **Biochemical Platform Conversion Targets**

	2007	2008	2009	2010	2011	2012 Target	2012 SOT
Minimum Ethanol Selling Price (\$/gal)	\$3.64	\$3.57	\$3.18	\$2.77	\$2.56	\$2.15	\$2.15
Feedstock Contribution (\$/gal)	\$1.12	\$1.04	\$0.95	\$0.82	\$0.76	\$0.74	\$0.83
Conversion Contribution (\$/gal)	\$2.52	\$2.52	\$2.24	\$1.95	\$1.80	\$1.41	\$1.32
Yield (Gallon/dry ton)	69	70	73	75	78	79	71
Feedstock							
Feedstock Cost (\$/dry ton)	\$77.20	\$72.90	\$69.65	\$61.30	\$59.60	\$58.50	\$58.50
Pretreatment							
Solids Loading (wt%)	30%	30%	30%	30%	30%	30%	30%
Xylan to Xylose (including enzymatic)	75%	75%	84%	85%	88%	90%	81%
Xylan to Degradation Products	13%	11%	6%	8%	5%	5%	5%
Conditioning							
Ammonia Loading (g per L hydrolysate liquor)	12.9	12.9	9.8	4.8	3.8	4.8	1.6
Hydrolysate solid-liquid separation	Yes	Yes	Yes	Yes	Yes	No	No
Xylose Sugar Loss	2%	2%	2%	2%	1%	1%	0%
Glucose Sugar Loss	1%	1%	1%	1%	1%	0%	0%
Enzymes							
Enzyme Contribution (\$/gal EtOH)	\$0.39	\$0.38	\$0.36	\$0.36	\$0.34	\$0.34	\$0.36
Enzymatic Hydrolysis & Fermentation							
Total Solids Loading (wt%)	20%	20%	20%	17.5%	17.5%	20%	20%
Saccharification Mode	Washed- solids	Washed- solids	Washed- solids	Washed- solids	Washed- solids	Whole- slurry	Whole- slurry
Combined Sacchrifcatn & Fermentatn Time (d)	7	7	7	5	5	5	5
Corn Steep Liquor Loading (wt%)	1%	1%	1%	1%	0.25%	0.25%	0.25%
Overall Cellulose to Ethanol	86%	86%	84%	86%	89%	86%	74%
Xylose to Ethanol	76%	80%	82%	79%	85%	85%	<mark>93</mark> %
Arabinose to Ethanol	0%	0%	51%	68%	47%	85%	54%

- Lower-severity pretreatment conditions (in conjunction with feedstock deacetylation) were demonstrated in continuous pilot scale reactors
- Higher enzymatic digestibilities of pretreated corn stover and ethanol yields from xylose enabled cost target achievement
- Strong collaboration with Biochemical Processing Interface Task enabled scale-up of deacetylation and pretreatment unit operations for the 2012 cellulosic ethanol demonstration runs in the 1 ton/day continuous pretreatment reactor



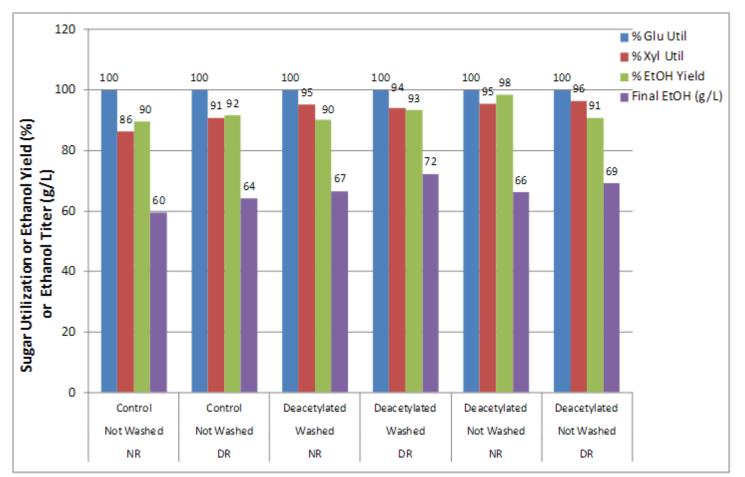
#### 200 kg/day Continuous Horizontal Reactor



#### 1 ton/day Continuous Horizontal Reactor

# Effect of Inhibitor Removal on Ethanol Production

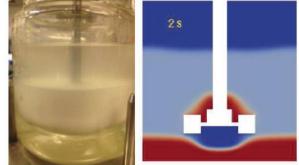
 Deacetylation with lower pretreatment temperature/acid loading allows for more complete xylose utilization and higher final ethanol titers of ~70 g/L (at 19.2% TS loading)

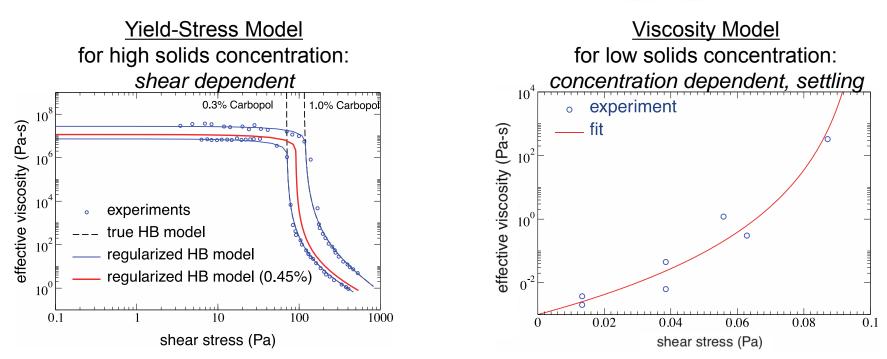


## Computational Fluid Dynamics of Biomass Slurries

#### Process Science of Enzymatic Hydrolysis

- Two CFD approaches to predict rheological behavior of enzymatically hydrolyzing biomass slurries in scaled biorefinery equipment
- Enzymatic hydrolysis reactors, fermenters, pumps, pipelines, and conveyors are some applications





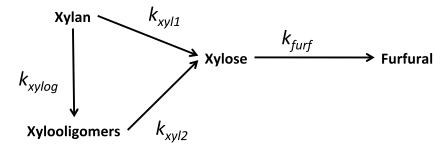
FY12 D-Milestone, "Implement coupled computational fluid dynamics and reaction kinetics simulations for the enzymatic hydrolysis of cellulose at dilute to moderate solids concentrations and validate against appropriate experimental data."

National Renewable Energy Laboratory

Innovation for Our Energy Future

## **Multi-Physics Pretreatment Model**

- Developing a model coupling mass transfer, heat transfer, phase transition, and reaction kinetics
- Literature indicates no other existing pretreatment model includes all these phenomena in one model



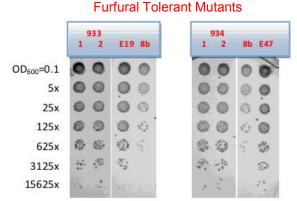
	Mass	Chemical	Heat	Phase
Model	Transfer	Reaction	Transfer	Transition
Kazi et al. (1997)	√(D)	×	×	X
Kim and Lee (2002)	√(D)	×	×	×
Jacobson and Banerjee (2006)	√(D)	×	×	×
Gustafson et al. (1983)	√(D)	$\checkmark$	×	×
Costanza et al. (2001)	√(D)	$\checkmark$	×	×
Mittal et al. (2009)	√(C)	$\checkmark$	×	×
Griggs and Kuhn (2011)	√(C)	$\checkmark$	×	×
Abasaeed et al. (1991)	×	$\checkmark$	$\checkmark$	×
Xu et al. (2010)	√(C)	×	$\checkmark$	$\checkmark$

FY13 E-Milestone, "Evaluate various mathematical modeling approaches for dilute-acid pretreatment of lignocellulosic biomass and select one that includes the most relevant chemical and physical mechanisms."

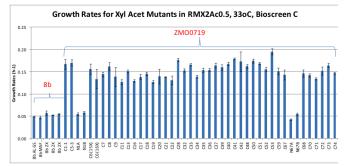
NATIONAL RENEWABLE ENERGY LABORATORY

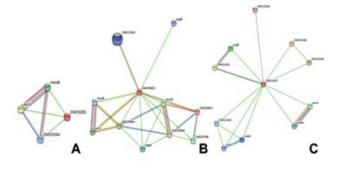
Genes related to toxicity in Zymomonas mobilis

- Identified and tested top candidate genes for resistance to furfural and acetate through both screening and selection of mutant library, and transcriptomics study
  - 3 genes conferring advantages over parental strains to acetate tolerance using xylose substrate and 6 genes related to improved furfural resistance
- Developed and improved synthetic genomics tools: NGS, RNA-seq, knockin-knockout gene testing methods and regulatory promoter development.
- Evaluated hydrolysate toxicity to support Xylose Yield subtask's D-milestone
  - Discovered new inhibitors in the hydrolysate work in progress for confirmation.



#### 3 g/L Furfural RMG Plate

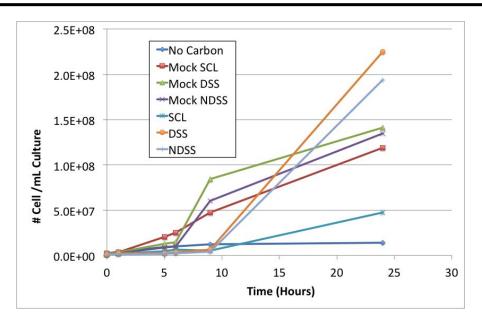


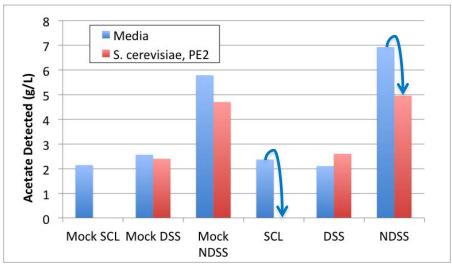


## Acetate Utilization

#### **Biological Toxicity Characterization & Mitigation**

- Assess possibility of utilizing biomass deacetylation liquor with acetate-assimilating hydrocarbon producing organisms
  - Test 8 microbial strains capable of producing hydrocarbon via variety of pathways
  - Evaluate growth in acetate w/ or w/o sugars and in spent caustic liquor and saccharified slurries
  - Saccharomyces cerevisiae PE-2 identified as most robust strain
    - Growth and acetate utilization in medium supplemented with either liquor or slurry.
- Oleaginous yeast and algae also show promise for acetate utilization



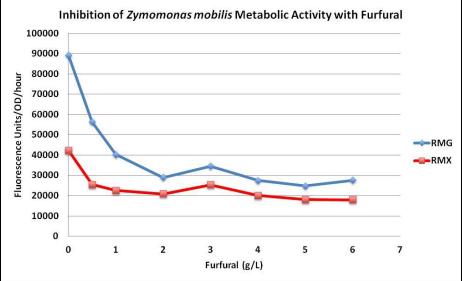


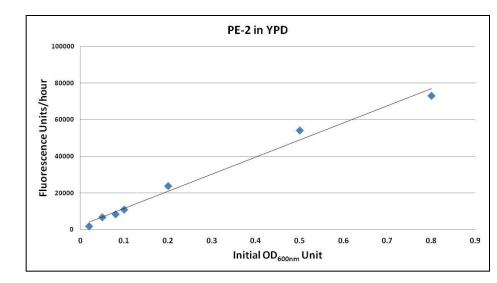
2013 E-Milestone. Assess the possibility of utilizing acetate in hydrolysate/slurry along with sugars or pre-hydrolysate liquor from biomass deacetylation with hydrocarbon producing organisms

NATIONAL RENEWABLE ENERGY LABORATORY

## **Inhibitor Identification**

- Identify inhibitors and their impact on model hydrocarbon fuel and/or intermediate-producing microorganisms in hydrolysate liquor and/or saccharified slurry from dilute acid pretreatment
- Develop high throughput assay that allows rapid measurement of toxic response for yeast and bacteria, aerobes and anaerobes
- Identify the top three toxicity contributors for 3 model hydrocarbon fuel and/or intermediate-producing microorganisms in a process relevant hydrolyzate and/or saccharified slurry
- Development of Alamar Blue fluorescent microtitre plate assay is nearly complete





## Furans to hydrocarbons

#### **Chemical Transformation**

- Synthesis of furan oligomers to C10 – C20 units
- HDO of furan oligomers to C10 C20 hydrocarbons (gasoline, diesel, jet fuel)
- Develop fuel pathways using furans from dilute acid pretreatment flash condensate
- Develop improved HDO catalysts
  - High activity (bimetallic nanocatalysts)
  - Hydrothermally stable
  - Acid-resistant (e.g. carboxylic acids)

